

THE COMPOSITION OF EIGHT *ACACIA* GUM EXUDATES FROM THE SERIES *GUMMIFERAE* AND *VULGARES**

D. M. W. ANDERSON and J. G. K. FARQUHAR

Department of Chemistry, The University, Edinburgh, EH9 3JJ, U.K.

(Received 5 October 1978)

Key Word Index—*Acacia*; Leguminosae; gum exudates; protein; glucose; chemotaxonomy; hybrid.

Abstract—An analytical study has been made of gum specimens from *Acacia hebeclada*, *A. kirkii*, *A. newbournii* and *A. reficiens* (all of the series *Gummiferae*) and of *Acacia erubescens*, *A. fleckii*, *A. mellifera* ssp. *mellifera* and *A. mellifera* ssp. *detinens* (all of the series *Vulgares*). The data obtained give further support for the main chemotaxonomic differences between the *Gummiferae* and *Vulgares* species recorded previously. In addition, two of the species studied have exceptional features; the gum exudate from *A. hebeclada* contains 9.4% of nitrogen; that from *A. erubescens* contains 12% of glucose.

INTRODUCTION

The chemical compositions of the gum exudates from nearly 90 *Acacia* species are now known; the names of the species concerned have been listed recently [2] in series according to Bentham [3] under the subgenera of Vassal [4]. In continuation of the search in this laboratory for *Acacia* gums with unusual chemical features worthy of full structural investigation in the future, this report presents the analytical data obtained for eight *Acacia* species whose gum exudates had not been studied

previously. In addition to revealing the existence of *Acacia* gums containing glucose and unusually high nitrogen contents, the analytical data obtained from this form of phytochemical survey are useful for chemotaxonomic purposes.

RESULTS AND DISCUSSION

The analytical data obtained for the eight species studied are shown in Table 1. The four species belonging to the *Gummiferae* have highly positive specific optical rotations; the species belonging to the series *Vulgares* have strongly negative rotations. This substantiates a

* Part 55 of the series "Studies of Uronic Acid Materials". For Part 54, see ref. [1].

Table 1. Analytical data for *Acacia* gums*

| | <i>Gummiferae</i> | | | | <i>Vulgares</i> | | | |
|--|-------------------------|-------------------------|--------------------------|----------------------|-----------------------|--------------------------|--|--|
| | <i>Acacia hebeclada</i> | <i>Acacia reficiens</i> | <i>Acacia newbournii</i> | <i>Acacia kirkii</i> | <i>Acacia fleckii</i> | <i>Acacia erubescens</i> | <i>Acacia mellifera</i> ssp. <i>detinens</i> | <i>Acacia mellifera</i> (Vahl) Benth. (Sudanese) |
| Moisture (%) | 12.5 | 11.7 | 13.0 | 10.8 | 12.5 | 13.2 | 12.1 | 8.7 |
| Ash (%) | n.d. | 2.4 | 4.2 | 1.4 | 4.0 | 3.9 | 3.6 | 2.9 |
| Nitrogen (%) | 9.4 | 0.65 | 0.14 | 0.09 | 0.58 | 1.08 | 1.3 | 1.45 |
| Hence protein (%) (N × 6.25) | 59 | 4.1 | 0.88 | 0.56 | 3.6 | 6.8 | 8.1 | 9.1 |
| Methoxyl (%) | 2.5 | 1.7 | 0.50 | 0.93 | 0.47 | 1.4 | 0.82 | 1.7 |
| Specific rotation, $[\alpha]_D$, degrees | +28 | +89 | +43 | +54 | −32 | −31 | −45 | −56 |
| Intrinsic viscosity, $[\eta]$, ml g ^{−1} | 13 | 12 | 13 | 8 | 13 | 8 | 21 | 23.5 |
| Molecular weight, MW × 10 ⁵ | n.d. | 3.77 | 3.65 | 2.08 | 4.15 | 2.0 | 10.4 | 4.1 |
| Equivalent weight | 521 | 1117 | 777 | 1817 | 918 | 874 | 822 | 843 |
| Hence uronic anhydride (%) | 33.8 | 15.8 | 22.6 | 9.7 | 19.2 | 20.1 | 21.4 | 20.9 |
| % Sugar composition after hydrolysis: | | | | | | | | |
| 4-O-Methylglucuronic acid | 15.0 | 10.1 | 3.0 | 5.6 | 2.8 | 8.4 | 4.9 | 10.2 |
| Glucuronic acid | 18.8 | 5.7 | 19.6 | 4.1 | 16.4 | 11.7 | 16.5 | 10.7 |
| Galactose | 44 | 41 | 45 | 36 | 39 | 39 | 44 | 43 |
| Glucose | — | — | — | — | 3 | 12 | — | — |
| Arabinose | 14 | 35 | 27 | 46 | 25 | 17 | 25 | 27 |
| Rhamnose | 8 | 8 | 7 | 8 | 14 | 12 | 9 | 9 |

* Collected in Namibia by Mr. Willy Giess, SWA Herbarium.

possible chemotaxonomic correlation that was noted previously [5].

The data presented for the two subspecies of *A. mellifera*, from widely differing geographical locations, indicate that the compositions of their gum exudates are extremely similar. The major difference recorded involves the MWs of the two specimens; this was also the major difference found in a recent study [1] of the variation between 15 different samples of *A. karroo* gum from widely different African locations. *Acacia laeta* has long been suspected to be a natural hybrid between *A. senegal* and *A. mellifera* [6]: the data now available for *A. mellifera* gum allow comparisons to be made with that published previously for the gums from *A. laeta* [7] and *A. senegal* [8]. The compositions of the three species are closely similar; it is interesting that, where differences occur, the values of the relevant parameters for *A. laeta* gum are intermediate between those for *A. mellifera* and those for *A. senegal*. This is further evidence that closely related *Acacia* ssp. give gum exudates that are closely similar in composition [1, 9, 10].

The gums from *A. fleckii* and *A. erubescens* have been found to contain glucose, as confirmed by the specific test involving glucose oxidase [11]. The presence of glucose in plant gums, first detected in the gum from *Anacardium occidentale* [11], may not be as uncommon as was at first supposed.

The most interesting feature of the analyses reported, however, concerns the remarkably high nitrogen content (9.4%) found for *A. hebeclada* gum. Until other very recent work, in which a nitrogen content of 7.2% was reported [12] for the gum from *A. torulosa* (*Juliflorae*), a nitrogen content of 1.66% [13] was the highest recorded for an *Acacia* gum. Previous studies [14] have shown the nitrogen content of the gums from *Acacia* and other genera to be proteinaceous in origin; attempts to free *Acacia* gum polysaccharides from proteinaceous matter without causing extensive degradation to the gum molecules were not successful [14]. It is important that the rôle played by the proteinaceous material in the production of the physico-chemical properties that are characteristic of the gum exudates should be clarified and that the existence of any direct polysaccharide-protein covalent linkage should be investigated. The relatively small protein content (<10%) in the *Acacia* gums studied previously did not make such experiments particularly attractive, but knowledge of the existence of *Acacia* gums containing 7–9% of nitrogen, indicative of a possible protein or polypeptide content of 40–55%, now makes the study of such materials a matter of urgency.

EXPERIMENTAL

Origin of gum specimens. Gum exudates from the following species belonging to the series *Gummiferae* were collected on

6 September, 1975, at Otjitambi, District Outjo, Namibia, by Mr. W. Giess (SWA Herbarium, Windhoek): *Acacia hebeclada* DC., *A. reficiens* Wawra and *A. newbournii* Burt Davy; gum from *A. kirkii* Oliver was collected by Mr. Giess on 4 September, 1975, at Otjovasandu, District Outjo. Gum exudates from the following species belonging to the series *Vulgares* were collected as follows: *A. fleckii* Schinz and *A. erubescens* Welw. ex Oliver by Mr. H. D. von Alvensleben at Kumkauas, District Grootfontein, on 29 September, 1975; *A. mellifera* ssp. *detinens* (Burch.) Brenan by Mr. W. Giess at Otjitambi, District Outjo, on 6 September, 1975; and *A. mellifera* (Vahl.) Benth. ssp. *mellifera* Brenan at Gardud Forest Reserve, Republic of the Sudan, on 20 March, 1978, by Mr. A. G. Seif-el-Din, Gum Research Officer, Republic of the Sudan.

Analytical methods. The standard analytical methods have been described [15]. The quantities of gum available for analysis were small; the amount available from *A. erubescens* did not allow determinations of ash nor MW to be made. The extraction of glucose, and its specific identification by means of glucose oxidase, has been described [11].

Acknowledgements—We are grateful to Rowntree-Mackintosh Ltd. (York) for financial support, and thank Mr. W. Giess and Mr. von Alvensleben, Windhoek (via Dr. Annabel Schreiber, Munich) and Mr. Seif-el-Din for the collection and identification of gum specimens.

REFERENCES

1. Anderson, D. M. W. and Pinto, G. (1979) *J. Linn. Soc. (London)* accepted for publication.
2. Anderson, D. M. W. (1978) *Kew Bulletin* 32, 529.
3. Benthams, G. (1875) *Trans. Linn. Soc. (London)* 30, 335.
4. Vassal, J. (1972) *Bull. Soc. Hist. Nat. Toulouse* 108, 1.
5. Anderson, D. M. W. and Dea, I. C. M. (1969) *Phytochemistry* 8, 167.
6. Anderson, D. M. W. and Smith, R. N. (1967) *Carbohydr. Res.* 4, 55.
7. Anderson, D. M. W., Dea, I. C. M. and Smith, R. N. (1968) *Carbohydr. Res.* 7, 320.
8. Anderson, D. M. W., Dea, I. C. M., Karamalla, K. A. and Smith, J. F. (1968) *Carbohydr. Res.* 6, 97.
9. Anderson, D. M. W. and Brenan, J. P. M. (1975) *Boissiera* 24, 307.
10. Anderson, D. M. W. and Bell, P. C. (1976) *Phytochemistry* 15, 301.
11. Anderson, D. M. W., Bell, P. C. and Millar, J. R. A. (1974) *Phytochemistry* 13, 2189.
12. Anderson, D. M. W., Farquhar, J. G. K. and Gill, M. C. L. (1979) *J. Linn. Soc. (London)* accepted for publication.
13. Anderson, D. M. W. and Gill, M. C. L. (1975) *Phytochemistry* 14, 739.
14. Anderson, D. M. W., Hendrie, A. and Munro, A. C. (1972) *Phytochemistry* 11, 733.
15. Anderson, D. M. W., Bell, P. C. and McNab, C. G. A. (1972) *Phytochemistry* 11, 1751.